Amendments to the Specification:

Please replace the paragraph beginning at page 7, line 19, with the following rewritten paragraph:

A LC-ESI-MS analysis of the reduced and pyridylethylated extract was performed, as may be seen from FIG. 2. To this end, reduced and pyridylethylated GndHCl extracts of CAP were injected onto reverse phase HPLC columns [~~Bio-Rad~~ BIO-RAD® HRLC series 800 system; columns C4 and C8 from Aquapore RP butyl (7 μm, 4.6x220 mm), Aquapore RP 300 (7 μm, 4.6x220 mm), Perkin-Elmer; Spherisorb 80-5C8 (220x4 mm); Marchery Nagel and Vydac protein C4 (4.6x220 mm)) pre-equilibrated with solvent A (0.1% v/v TFA in water) and eluted with a linear gradient of increasing concentration of solvent B (80% v/v acetonitrile and 0.1% v/v TFA): 0-15% B in 5 min, 15-27% B in 40 min, 27-35% B in 2 min, isocratic at 35% B for 3 min, 35-43% B in 25 min, 43-56% B in 50 min, 56-70% B in 5 min, 70-100% B in 10 min and isocratic at 100% B for 5 min].

Please replace the paragraph beginning at page 8, line 4, with the following rewritten paragraph:

Since the average Mw of the protein designated CSP14 could not be assigned with the above method, the peak fraction was dissolved in 500 μl of 25% solvent B (0.05% (v/v) TFA/80%, v/v ACN). For SDS-PAGE, a 10 μl aliquot was dried in ~~speedvac~~ a SPEEDVAC® low vacuum system and dissolved in 20 μl SDS-sample buffer and analyzed on gradient 10-20% T ready Tris-Tricine acrylamide gels using the miniprotean 3 system from ~~Bio-Rad~~ BIO-RAD®. Protein bands were visualized by staining the gels in the staining solution [0.5% (w/v) Commassie Brilliant Blue R250 in 30% (v/v) methanol and 10% (v/v) acetic acid] for 1 hr followed by destaining [30% (v/v) methanol plus 10% (v/v) acetic acid] until bands were visible against the clear background (Graffin, Methods Enzymol. 182 (1990) 425-477). Accordingly it could be observed that CSP 14 corresponds to a protein having a molecular weight of about 14 kDa.